

REVIEW

Open Access



Nuclear medicine imaging in non-seminomatous germ cell tumors: lessons learned from the past failures

Narjess Ayati^{1,2†}, Emran Askari^{3†}, Maryam Fotouhi⁴, Masume Soltanabadi⁵, Atena Aghaee³, Hesamoddin Roustaei⁶ and Andrew M. Scott^{7,8,9,10*}

Abstract

There is an unmet need for a more accurate molecular imaging radiotracer in the field of non-seminomatous germ cell tumors (NSGCT). The clinical problem is that no single imaging modality is able to differentiate teratoma from necrotic tissue in NSGCTs, which the nuclear medicine techniques are no exception. The exponential growth in the list of potentially promising radiotracers may hold promise in the future for imaging of NSGCTs. Here, we have reviewed the past efforts and potential future advances in this field.

Keywords Non-seminomatous germ cell tumor, Teratoma, Retroperitoneal residual mass, Radiotracer, Imaging

Epidemiology

Testicular cancer is the most common malignancy in men aged 15–45 years and accounts for 1–1.8% of all male cancers [1, 2]. Of testicular malignancies, 95% are germ cell tumors (GCTs) [3]. About 50% of patients with GCT present with advanced non-seminomatous germ cell tumors (NSGCT) [4, 5].

The imaging problem in the field of GCT

Following radical orchiectomy and adjuvant cisplatin-based triplet chemotherapy for the treatment of stage IIB–III NSGCT, there is a 30–40% chance of retroperitoneal mass persistence [6–9]. In 40–51% of these cases the retroperitoneal masses represent necrotic/fibrotic tissues, while 30–47% are teratomas, and the remaining 6–17% of cases are different histopathologies simply grouped as viable GCTs [10–12]. Post-chemotherapy retroperitoneal lymph node dissection (PC-RPLND) aims to eradicate all the remaining viable malignant tissue in the advanced NSGCTs, however distinguishing residual viable tumor from post-therapy changes remains a major challenge when deciding if surgery is required [13].

[†]Narjess Ayati MD and Emran Askari MD contributed equally to this work.

*Correspondence:

Andrew M. Scott
andrew.scott@austin.org.au

¹Department of Theranostics and Nuclear Medicine, St. Vincent's Hospital, Sydney, NSW, Australia

²St. Vincent's Clinical School, University of New South Wales, Sydney, NSW, Australia

³Nuclear Medicine Research Center, Mashhad University of Medical Sciences (MUMS), Mashhad, Iran

⁴Advanced Diagnostic and Interventional Radiology Research Center (ADIR), Cancer Institute, Imam Khomeini Hospital Complex, Tehran University of Medical Sciences, Tehran, Iran

⁵Rajaie Cardiovascular Medical and Research Center, Iran University of Medical Sciences, Tehran, Iran

⁶Department of Nuclear Medicine, University Hospital Salzburg, Paracelsus Medical University, Salzburg, Austria

⁷Tumour Targeting Laboratory, Olivia Newton-John Cancer Research Institute, Melbourne, VIC, Australia

⁸School of Cancer Medicine, La Trobe University, Melbourne, VIC, Australia

⁹Department of Molecular Imaging & Therapy, Austin Health, 145 Studley Road, Heidelberg, VIC 3084, Australia

¹⁰Department of Medicine, University of Melbourne, Melbourne, VIC, Australia



Table 1 Anatomic features for differentiation and prognostication of mature teratoma versus immature or growing teratoma across a prognosis range from poor to good [20, 22, 109]

Radiologic feature	Prognostic Impression	
	Poor	Good
Echogenicity	Solid	Cystic with heteroechoic and hyperechogenic foci ^{*1} ; onion ring appearance ^{*2}
Vascularity	Hypervascular	Hypovascular
Borders	Indistinct	Distinct
Contrast enhancement	Heterogeneous enhancement ^{*3}	Mild
Nodular formation	Yes	No
New lesions or increase in size of previous lesions	Frequent	Infrequent
Location	Retroperitoneal, mediastinal and intracranial	Confined to retroperitoneum
Spontaneous regression	Rare	Reported in burn-out teratomas

^{*1} Mature teratomas may eventually grow in previous sites of metastasis, presenting as cystic changes with heterogeneous density changes containing calcification and fat. In these cases, serial follow-up CT imaging may be indicated before proceeding to surgery

^{*2} Associated with dermoid cysts

^{*3} For contrast-enhanced CT, solid portion of the mass along with septations are enhanced while the cystic fat-containing component usually remains unchanged

Several studies have shown that not only the viable tumors but also those with teratoma are at increased risk of recurrence [14–18]. For instance, Nestler and colleagues [14] analyzed data from a multi-center cohort of 1204 non-seminomas who underwent PC-RPLND and observed a significantly increased risk of recurrence by five years in the viable GCT/teratoma subgroups compared to patients with only necrosis (81% and 59%, vs. 19%, respectively, $p < .001$). Moreover, teratomas should be resected due to their resistance to chemoradiation, compressive effect on adjacent organs, and their ability to undergo malignant transformation, especially in the subtype of teratoma with somatic malignancy [11, 19]. Therefore, differentiation of teratomas from necrosis/fibrosis is clinically relevant.

Retroperitoneal teratomas are usually asymptomatic, and tumor markers frequently fall within the normal range, except in cases of mixed GCT or those with mucinous or hepatoid differentiation. Therefore, its detection and follow-up are highly reliant on anatomic imaging [19, 20]. Mature teratomas, also referred to as differentiated teratomas, usually present as low attenuation retroperitoneal masses with less aggressive behavior [21]. There are some conventional imaging features that are useful for the differentiation of mature teratoma from immature or growing teratoma (Table 1). However, irrespective of

Table 2 Prognosticators of post-chemotherapy retroperitoneal residual mass

Favorable prognosis	Unfavorable prognosis
No teratoma component in the orchiectomy specimen [27] ^{*1, *2}	Abnormal tumor markers [53]
Normal pre-chemotherapy AFP and HCG [27]	Multiple FDG-avid residual masses [53]
Elevated pre-chemotherapy LDH [27] ^{*3}	Post-chemotherapy nodal size [24]
Small residual mass (< 10–20 mm in small transverse diameter) [27] ^{*2}	Supra-diaphragmatic lymph nodes and/or visceral metastasis [6]
Marked residual mass reduction (> 70–90%) [27]	Prior history of relapse [25]
< 10% residual viable cells in the PC-RPLND specimen [16]	Late-onset relapse (i.e., > 2 years) following chemotherapy [25]

^{*1} Predictors of teratoma in post-chemotherapy retroperitoneal lymph node dissection (PC-RPLND) specimen includes presence of teratoma and yolk sac tumor in the orchiectomy specimens [11]. Absence of teratoma in the orchiectomy specimen does not exclude the presence of teratoma in PC-RPLND [24, 25]

^{*2} Predictors of necrosis in PC-RPLND specimen includes presence of seminomatous and absence of teratomatous elements in the primary tumor, normal pre-chemotherapy beta-hCG and AFP levels, small pre-chemotherapy (cutoff: < 2 cm) or post-chemotherapy (cutoff: ≤ 1 cm) lymph nodes and > 50% mass size reduction following chemotherapy [26, 30]

^{*3} The International Germ Cell Cancer Collaborative Group prognostic classification also considers elevated LDH levels in the poor prognostic group [9]

their subtype, teratomas should be resected as per international guidelines [22, 23].

Review of the guidelines

Unfortunately, no non-invasive diagnostic modality or validated risk calculator can accurately determine the nature of the residual mass (Table 2) [6, 11, 13, 24–29]. Therefore, the EAU guidelines recommends resecting the post-chemotherapy residual mass if > 1 cm in greatest diameter on contrast-enhanced CT (ceCT) whenever feasible [30]. In this context, PC-RPLND serves as both a diagnostic and a therapeutic tool [28]. This approach will over-treat almost half of the patients while leaving 25% risk of teratoma and 5% risk of viable tumor in small sub-centimetric lesions, which have an overall 6–9% risk of relapse and may be captured by subsequent imaging [3, 11, 27]. In this context, surgery is often without oncological benefit [14] and major post-surgical complication rates are non-negligible according to a systematic review [31].

Some authors have suggested that the incorporation of non-invasive imaging modalities, such as ¹⁸F-FDG PET/CT, into the management algorithm may allow better prediction of viable residual tumors and, thus better risk stratification in this setting [4]. However, the NCCN guideline [32] recommends abdominopelvic ceCT, MRI, and CXR as modalities for imaging first- and second-line

chemotherapy patients and routine follow-up cases, which is also consistent with other guidelines (e.g., ESMO, SWENOTECA) [33–35]. The NCCN guidelines currently recommend against the routine use of ^{18}F -FDG PET/CT while considering its possible usefulness for surveillance of patients in the post-chemotherapy status [32].

Why ^{18}F -FDG PET/CT was not so successful?

The utility of ^{18}F -FDG PET/CT in patients with NSGCT has been a topic of considerable debate, with views ranging from some to no benefit [36–38]. Viable tumors have significantly higher FDG uptake (Fig. 1) as compared to the generally low FDG uptake in necrosis, fibrosis, or teratoma [39]. Also, a negative ^{18}F -FDG PET/CT scan has been linked with increased overall survival [40]. ^{18}F -FDG PET/CT has also been investigated in the context of relapse following definitive NSGCT treatment. It has also been shown that the levels of tumor markers (i.e., LDH, AFP, and hCG) have a significantly positive correlation with the ^{18}F -FDG uptake [41].

Contrary to the studies mentioned above, in one study, ^{18}F -FDG PET was even inferior to CT for differentiation of necrosis/fibrosis from teratoma [42] (Fig. 2), which is considered the main disadvantage of ^{18}F -FDG PET/CT [43–46]. Moreover, ^{18}F -FDG PET/CT may falsely show extensive uptake in post-chemotherapy inflammatory

changes [47], especially if imaged early post-treatment [12] (Fig. 3). Furthermore, it will miss small (i.e., <5–10 mm) lesions [48, 49], leading to high relapse rates among the PET-negative patients [50]. Therefore, its application for routine staging of NSGCT is discouraged since it will not have a clear added value to the standard ceCT and will not alter the treatment management [7, 23, 51, 52]. Also, for prediction of response to chemotherapy, ^{18}F -FDG PET/CT has not been shown to be superior to ceCT or serum tumor markers, although being a strong predictor of pathologic viable disease [37, 53, 54].

In one study in NSGCT patients examined 85 residual lesions, with 32 (38%) showing increased tracer uptake, resulting in a sensitivity of 59%, specificity of 92%, NPV of 62%, and PPV of 91% [53]. Therefore, ^{18}F -FDG PET positivity may be clinically relevant in evaluation of residual masses. In another study, quantitative ^{18}F -FDG PET analyses indicated significant differences between mature teratoma and necrosis or scar tissue, supporting its use for evaluating residual lesions post-chemotherapy [55]. Additionally, another study found that an SUV greater than 5 is more likely to be linked to viable GCT than necrosis, fibrosis, or mature teratoma [36]. However, the overall diagnostic benefit of ^{18}F -FDG PET/CT over traditional markers and CT scans for suspected NSGCT recurrence remains uncertain (Fig. 4), though patients

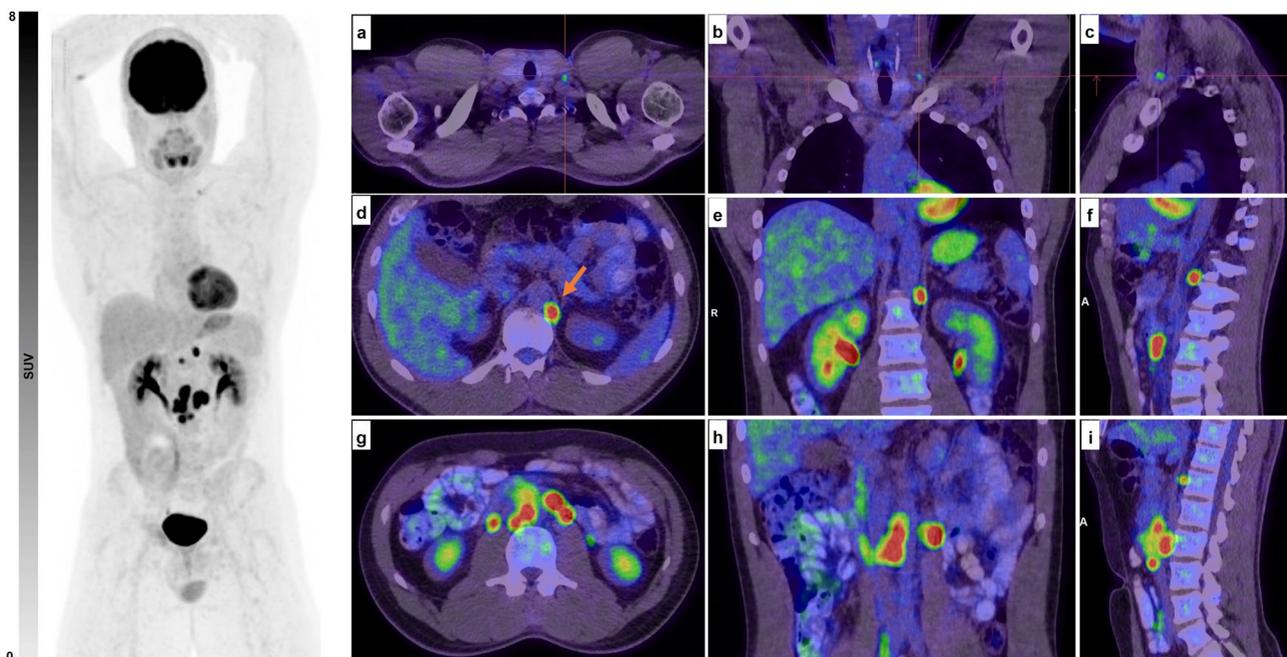


Fig. 1 A 32-year-old male with non-seminomatous testicular cancer, initially treated with orchidectomy, presented with suspicious para-aortic lymph nodes on CT and underwent an FDG PET scan. The images reveal a subcentimetre left supraclavicular lymph node (SUVmax 3.5; **images a-c, crosshairs**), bilateral intensely FDG-avid retrocrural lymph nodes (SUVmax 8 on the right and 12 on the left; **images d-f, red arrow**), and bilateral para-aortic lymphadenopathy extending from the axial level of L1 to L2/L3 on the right (SUVmax 21) and from the axial level of L1/L2 to L2/L3 on the left (SUVmax 25; **images g-i**). A subsequent biopsy of the left supraclavicular node confirmed metastatic involvement. Retroperitoneal lymph node dissection also confirmed multifocal retroperitoneal nodal metastases

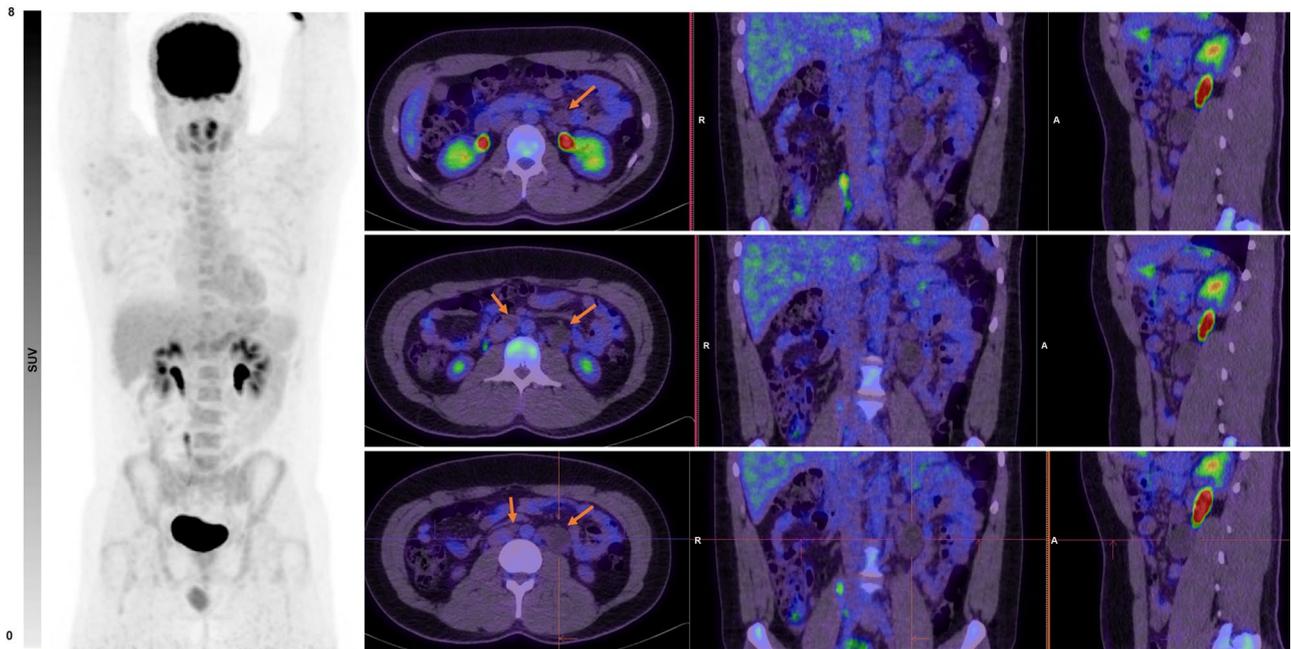


Fig. 2 A 30-year-old male with a non-seminoma germ cell tumor, previously treated with a left orchidectomy. The PET scan demonstrates multiple low-density nodal lesions in the retroperitoneum, including at the aortocaval and left para-aortic stations (**red arrows**), with no increased FDG uptake. The patient subsequently underwent retroperitoneal lymph node dissection, which revealed multiple nodal metastases

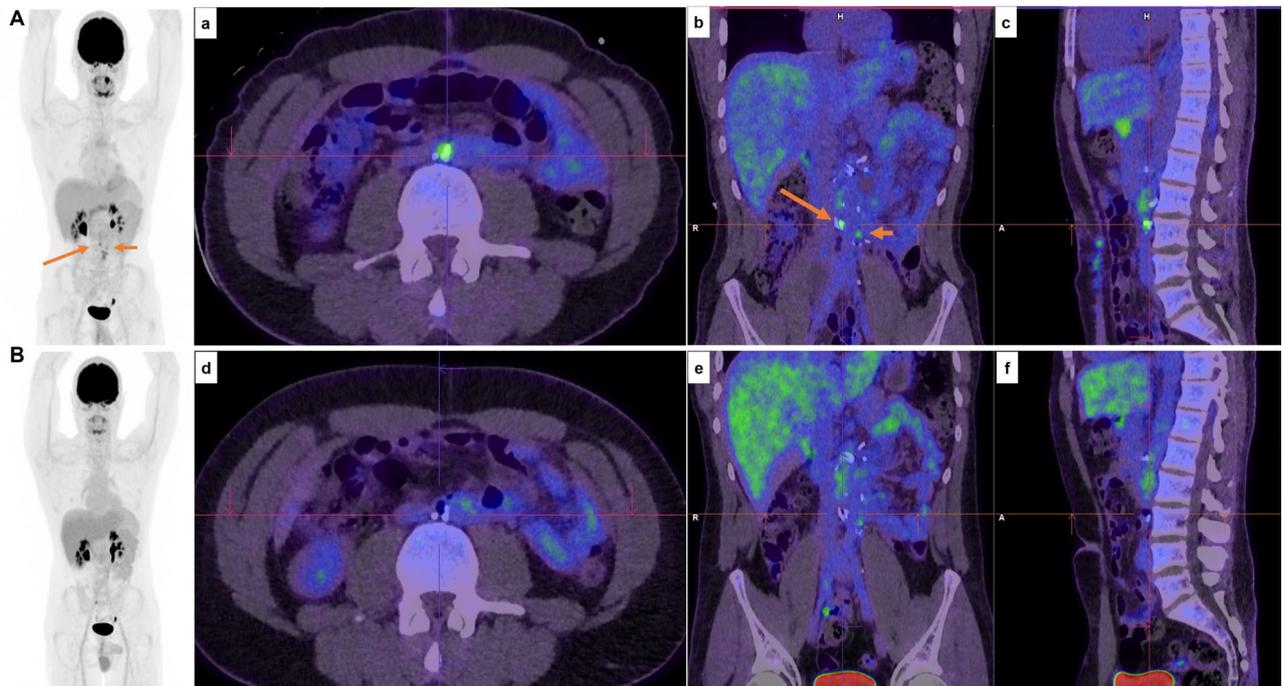


Fig. 3 A 33-year-old male with a history of non-seminoma germ cell tumor, previously treated with left orchidectomy, chemotherapy, and retroperitoneal lymph node dissection for bulky para-aortic lymph node metastases, presents for a progress assessment. The PET scan (**Image A**) demonstrates mild foci of uptake around the aortocaval (SUV max 3.9) and left para-aortic (SUV max 3.3) regions at the level of L3, adjacent to surgical clips (**red arrows, a-c**), which were reported as indeterminate (either post-surgical inflammatory changes or residual disease). Twelve months later, a follow-up PET scan (**Image B**) showed an interval reduction in the intensity of retroperitoneal foci of uptake (**d-f**), consistent with resolving post-operative inflammatory changes

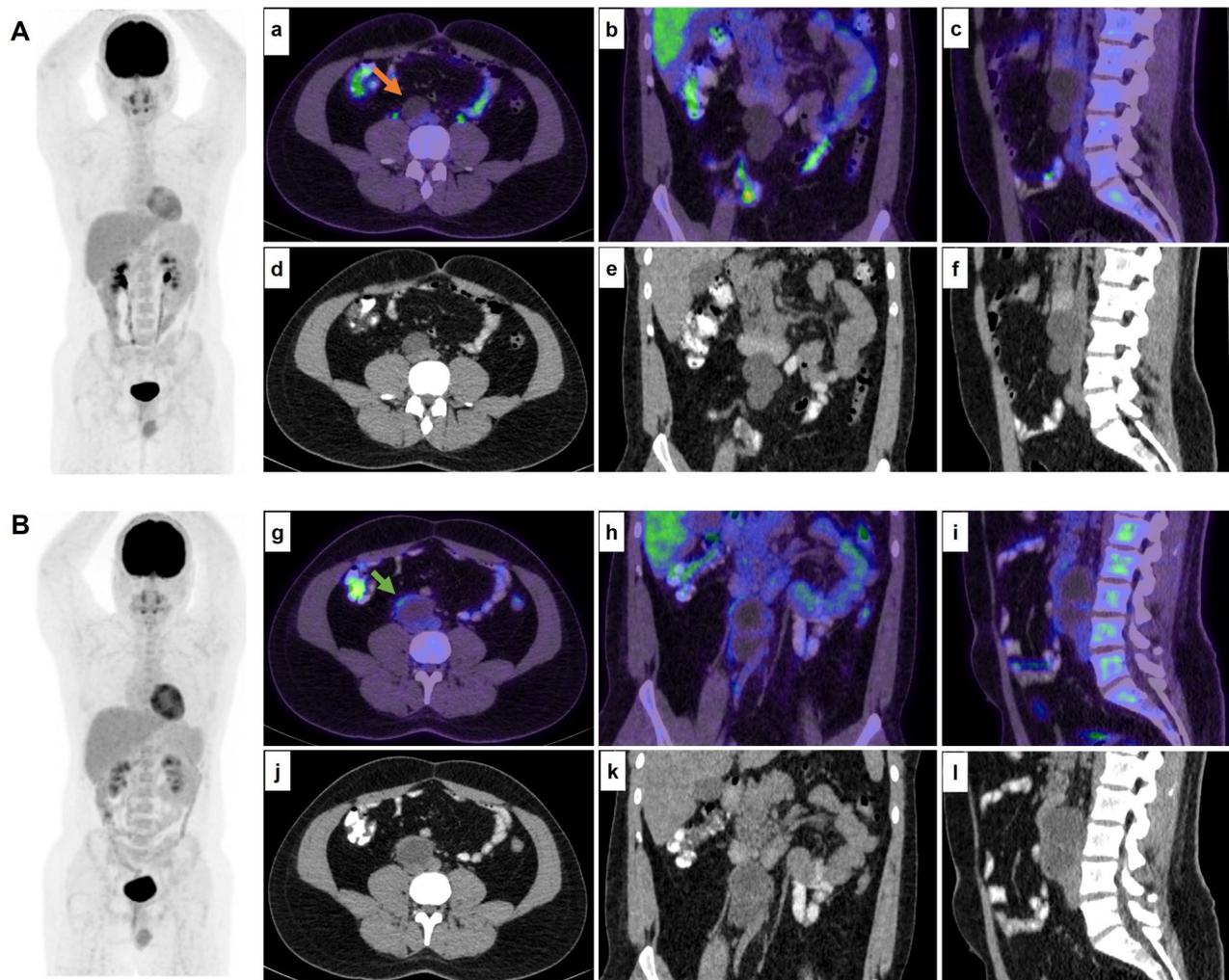


Fig. 4 A 23-year-old male with non-seminomatous testicular cancer, treated with orchidectomy followed by chemotherapy, presented with suspicious retroperitoneal lymphadenopathy on CT and underwent an FDG PET scan (**Image A**). The axial (**a, d**), coronal (**b, e**), and sagittal (**c, f**) views show enlarged hypoattenuating retroperitoneal lesions with no metabolic activity (**red arrow**). Serial CT scans demonstrated ongoing enlargement of these lesions. Ten months later, a follow-up PET scan (**Image B**) revealed further enlargement of the hypoattenuating retroperitoneal lesions with interval development of peripheral metabolic activity (**images g-l, green arrow**), highly suggestive of nodal metastases. Subsequent nodal dissection confirmed the presence of nodal metastases on histopathology

with elevated tumor markers with equivocal CT findings might benefit from ^{18}F -FDG PET/CT [56, 57].

In conclusion, in the era of “forget the PET” approach, ^{18}F -FDG PET/CT may infrequently be requested for rising tumor markers with normal ceCT and those with equivocal ceCT findings [58, 59]. Kinetic analysis may improve its diagnostic performance but is not performed in routine clinical practice, and has not been fully validated in large prospective studies [55].

Teratoma imaging with other radiopharmaceuticals: a land of failures

The multi-layered embryologic origin of teratoma sometimes contains immature neuro-ectodermal elements, which may eventually trap radioiodine or radiotracers targeting the somatostatin-receptors [60, 61]. Multiple

research groups have explored the added value of other PET radiopharmaceuticals, most of which were not very successful (Table 3). Perhaps the best one was imaging with radiopharmaceuticals targeting integrins. Yet, none of these radiotracers find their way into clinical practice. A more in-depth review of the experience gained by each imaging modality is discussed as follows.

PET tracers

^{11}C -tyrosine PET

A study showed that ^{11}C -tyrosine is not suited to visualize the apparently slowly proliferating NSGCT or to determine the nature of a residual mass after chemotherapy [62].

Table 3 Teratoma imaging with non-¹⁸F-FDG radiotracers *1

Radiotracer class [Name of the radiotracer]	Research aim	Sample size	Study type	Encouraging results?	First author (Reference)
Integrins - α _v β ₃ [^{99m} Tc-HYNIC-RGD]	Mature teratoma vs. necrosis	20 rats	Preclinical animal study	Yes, for α _v β ₃ imaging No, for FDG	Aide [110]
Integrins - α _v β ₃ [^{99m} Tc-3PRGD2]	Detection of hiPSC-derived teratoma	4 rats	Preclinical animal study	Yes, for α _v β ₃ imaging No, for FDG	Li [67]
Integrins α _v β ₃ [⁶⁴ Cu-DOTA-RGD4]	In vivo visualization of teratoma formation	12 rats	Preclinical animal study	Yes, for α _v β ₃ imaging No, for FDG and FLT	Cao [111]
Integrins - α _v β ₃ [^{99m} Tc-HYNIC-RGD]	Biodistribution and imaging of α _v β ₃ -negative and positive tumors	4 rats	Preclinical animal study	Equivocal, no definite conclusion regarding the main reason for integrin uptake (i.e., vasculature vs. cellular expression)	Bohn [112]
Radioiodine - ¹²⁴ I	In vivo visualization of teratoma formation	9 rats	Preclinical animal study	Yes, highly specific; Tracer uptake correlated with teratoma weight and washed out with perchlorate	Lehner [113]
¹⁸ F-FLT [39-deoxy-39–18 F-fluorothymidine]	Added value of FLT on top of FDG PET	11 patients (2 teratoma cases)	Case series	No, FDG and FLT were both falsely negative.	Pfannen-berg [63]
⁶⁷ Ga-Citrate	Detection rates in various GU malignancies	11 teratoma cases (16 lesions)	Retrospective clinical cases	No, low detection rate (25%)	Sauerbrunn [72]
PSMA: - ⁶⁸ Ga-PSMA/ ¹⁷⁷ Lu-PSMA	Single dose of ¹⁷⁷ Lu-PSMA RLT in a case of refractory mixed GCT	1 patient	Case report	No, rise in AFP and evidence of tumoral progression in the hepatic lesion despite sufficient PSMA uptake	Simsek [66] *2
FAPi: - ⁶⁸ Ga-FAPi-04	Comparison with FDG uptake	1 patient	Case report	Equivocal, FAPi (moderate uptake) vs. FDG (low uptake)	Kaplan [1] *3
¹⁸ F-Fluciclovine	Detection of residual disease/teratoma	10 patients (5 teratoma cases)	Case series	No, low detection rate (40%) in the teratoma subgroup	Woldu [64]

*1 Staging with somatostatin receptor scintigraphy and mIBG has been attempted for teratoma cases associated with TNET. Due to its rarity, the added value of these imaging modalities to conventional imaging is not yet demonstrated [114, 115]. Moreover, there are a few case reports/series for incidental detection of non-seminomatous GCT with ^{99m}Tc-pertechnetate and ^{99m}Tc-MDP or non-visualization of teratocarcinoma using ²⁰¹Tl [71–73]

*2 Tried in the light of prior human tissue studies [65]

*3 Phase I studies are ongoing [116]

GCT=Germ cell tumor; GU=Genitourinary; hiPSC=human-induced pluripotent stem cell; RLT=Radioligand therapy; TNET=Testicular neuroendocrine tumor

¹⁸F- fluorothymidine (¹⁸F-FLT)

In a small series of 11 patients (10 NSGCT, 1 seminoma) with metastatic NSGCTs, Pfannenberget al. compared the diagnostic value of ¹⁸F-FLT, which measures tumor cell proliferation, with ¹⁸F-FDG PET/CT. Despite the lower incidence of false-positive results with ¹⁸F-FLT PET than with ¹⁸F-FDG PET, the low negative predictive value of ¹⁸F-FDG PET could not be improved by the application of the ¹⁸F-FLT (60% and 50%, respectively). Therefore, PET-negative residual masses after chemotherapy of metastatic NSGCT still require resection. The low sensitivity of ¹⁸F-FLT PET/CT for the detection of viable residual tumors in this study may be related to the lower tissue uptake of ¹⁸F-FLT than of ¹⁸F-FDG in GCTs. Positive results on ¹⁸F-FDG PET after chemotherapy correlated strongly with the presence of viable tumors. For prediction of response after completion of chemotherapy, the final PET/CT scan, whether performed using ¹⁸F-FDG or using ¹⁸F-FLT, cannot be replaced by early response evaluation [63].

¹⁸F-fluciclovine

The potential use of ¹⁸F-fluciclovine for molecular imaging of NSGCTs was evaluated in a small prospective study, which revealed poor sensitivity and specificity in detecting teratoma from fibrosis/necrosis in patients with residual masses undergoing PC-RPLND. Half of the negative ¹⁸F-fluciclovine PET/CT cases were found to have residual disease/teratoma following surgery. The low utility of ¹⁸F-fluciclovine PET/CT in guiding the management of NSGCT post-chemotherapy was evident, with sensitivity and specificity rates at 29% and 33%, respectively [64].

Prostate-Specific Membrane Antigen (PSMA)

Prior human tissue studies have shown the expression of PSMA in some cases of NSGCT [65]. A case report of metastatic mixed (immature teratoma and yolk sac carcinoma) testicular GCT with acceptable tumor-to-background ratio was treated with the therapeutic

counterpart of PSMA, namely ^{177}Lu -PSMA, was not successful [66].

Integrins

Since their discovery in 2006, induced pluripotent stem cells (iPSCs) have gained increasing interest in tissue regeneration and transplantation therapies. However, teratoma formation after iPSC transplantation is one of the most serious drawbacks of this procedure. In a study, it was investigated whether human iPSC-derived teratomas could be detected by an integrin-targeting agent, $^{99\text{m}}\text{Tc}$ -PEG4-E[PEG4-c(RGDfK)]2 ($^{99\text{m}}\text{Tc}$ -3PRGD2). Gamma camera imaging with $^{99\text{m}}\text{Tc}$ -3PRGD2 may be a promising approach for the non-invasive monitoring of tumorigenicity after human iPSCs transplantation [67]. Unfortunately, these preclinical observations were never explored on human subjects.

Non-PET Tracers

SPECT/CT's spatial resolution is challenged over PET/CT, particularly in small lesions that are not always metabolically active, including NSGCTs [68]. This gap can be bridged with advanced quantification and reconstruction techniques and multi-pinhole collimators focusing on gamma rays [69]. PET facilities are preferred over SPECT in regions where both options are available; however, SPECT remains cost-effective in specific clinical applications and resource-limited countries [70, 71]. The results of teratoma imaging utilizing SPECT are unsatisfactory [71–80]. Small case series and case reports reported non-visualization of teratocarcinoma using Tl-201 [71] or incidental detection of NSGCTs using $^{99\text{m}}\text{Tc}$ -MDP due to ossification and cartilage tissue in teratoma [72], and $^{99\text{m}}\text{Tc}$ -pertechnetate due to increased flow in tumoral tissue [73].

Gallium-67

Traditionally, ^{67}Ga scintigraphy was considered valuable in assessing the intra-abdominal spread of malignant tumors of the testes. However, it appeared that metastatic tumors of the embryonal-cell and seminoma type, compared to teratoma, are more readily detectable by gallium-67 scanning [72]. Although its application for imaging of NSGCT were disappointing and discontinued [73, 74], its utility for staging in seminoma also became obsolete after the introduction of ^{18}F -FDG PET/CT [75].

Radiolabeled antibody

Radioimmunodetection captures tumor-specific or tumor-associated markers by preferentially accumulating tumor-specific antisera in tumoral tissues. Murine teratocarcinomas were localized using external gamma-ray scintigraphy with ^{131}I -labeled monoclonal antibodies. By removing background radioactivity from the control

monoclonal antibody 123 of the same immunoglobulin class, detection was enhanced [76]. Javadpour et al. utilized ^{131}I -labeled antibodies targeting tumor-associated antigens in testicular cancer to identify occult disease [77]. The limited sensitivity of this approach in identifying lesions under 2×2 cm and interference from background radioactivity limit its practical applicability. Epenetos et al. investigated placental alkaline phosphatase-targeting indium-111 monoclonal antibodies. Their study showed improved ovarian, cervical, and testicular cancer diagnosis. However, there are still ongoing issues regarding the pharmacokinetics and immunogenicity of antibodies, despite the positive outcomes shown [78].

Potential of imaging teratoma with novel radiotracers

Fibroblast activation protein inhibitor (FAPi)

FAP-targeting PET tracers have been extensively studied in both malignant and non-malignant entities [79, 80]. FAPi ligands may have a complementary role in detecting metastatic lymph nodes, especially if coupled with ^{18}F -FDG PET imaging in various cancers [81]. In addition, FAPi PET imaging has been shown to be able to detect fibrotic tissue in various scenarios (e.g., post-chemotherapy fibrosis in GI malignancies, idiopathic retroperitoneal fibrosis, and various non-malignant fibrotic pathologies) [79, 82]. However, to the best of our knowledge, till today, there has been no published comprehensive paper on FAPi PET tracers in NSGCTs.

Regarding FAP application in detecting teratoma, there is insufficient data in the literature. Xi et al., in a study conducted to compare ^{68}Ga -FAPi-04 PET/MR and ^{18}F -FDG PET/CT in ovarian tumors in 2023, and reported that of all the included cases, two patients had teratoma (one considered benign pathology and the latter borderline) [83]. In a study comparing ^{68}Ga -FAP-2286 PET/CT head-to-head with ^{18}F -FDG PET/CT in various malignancies, one case of metastatic yolk sac germ cell tumor was evaluated and showed better LN detection performance of FAPi ligand over ^{18}F -FDG with a better target-to-background ratio (separate values are not reported). Visually, in comparison to [^{68}Ga]Ga-FAP-46, this new FAP ligand seems to have a higher uptake [84].

There are two additional papers that each report one case of GCT imaged with FAPi radiotracers. Dai et al. [85] reported a rare case of extragonadal yolk-sac tumor in which ^{68}Ga -FAPi PET/MR outperformed ^{18}F -FDG PET/CT in the detection of the cranial lesion. The other case report looked at ^{68}Ga -FAPi-04 PET/CT and ^{18}F -FDG PET/CT in a person who had mixed testicular GCT that was 65% post-pubertal teratoma, 25% yolk sac and 10% seminoma. The retroperitoneal and lung nodules showed a slight uptake of FDG. Meanwhile, FAPi imaging revealed a mild-moderate uptake in the affected lesions (SUV_{max} of 3.9) [1].

CXCR4

CXCR4 is a seven transmembrane domain G protein-coupled receptor (GPCR) that contributes to chemotaxis, invasion, angiogenesis, aggressiveness, tumor progression, proliferation, and metastasis [86]. The CXCR4 ligand is frequently overexpressed in various types of cancer [87]. The CXCL12/CXCR4 pathway has a confirmed and significant role in the adult human testis microenvironment and is also expressed in gonadal and extragonadal GCTs [88, 89]. Yet, in all the publications concerning cellular studies, there is no clinical data to support this hypothesis.

Ghrelin

Testicular tumors differentially express the Ghrelin receptor, a GPCR involved in growth hormone secretion and food intake. In order to enhance in vivo stability and incorporate the Fluorine-18 isotope for PET imaging, the ghrelin ligand has recently undergone some modifications. This novel PET agent has been shown to have a high affinity for the ghrelin receptor in biochemical and preclinical studies [90, 91]. For now, there is no available clinical (in humans) data regarding Ghrelin receptor imaging in NSGCT.

Other solutions beyond nuclear medicine approaches

Magnetic Resonance Imaging

Advancements in the field of MRI in patients with testicular cancer are threefold. First, follow-up whole-body MRI may be employed in the future in lieu of ceCT due to concerns regarding radiation exposure and in the light of positive non-inferiority trials recently published in this regard [92, 93]. Second, new MRI sequences, namely T₁-Dixon and T₂-BLADE, have been shown to propose better performance in detecting retroperitoneal metastasis and were better than DWI-MRI in a prospective study [94]. Third, lymphotropic nanoparticle MRI (LNMRI) utilizes nanoparticles that aggregate with a mixed signal within pathological nodal tissue [95]. Its use is superior to conventional MRI, according to meta-analyses, in terms of both sensitivity (88% vs. 63%) and specificity (96% vs. 95%) [96, 97]. Harisinghani et al. [97] conducted a pilot trial of LNMRI to detect occult metastases in 18 men with testicular cancer. LNMRI had improved sensitivity (88% vs. 71%) and specificity (92% vs. 68%) compared to MRI or CT size criteria among these patients. Likewise, LNMRI was 100% sensitive in detecting positive lymph nodes less than 10 mm, that would not have been considered suspicious on conventional imaging. However, the lengthy duration (24–36 h) between nanoparticle injection and MRI, along with the requirement for an experienced radiologist to accurately interpret the images, restricts its adoption [54].

The potential role of radiomics

Literature regarding the added value of radiomics to differentiate necrosis/fibrosis from teratoma is emerging, with some studies showing encouraging results [98, 99]. As of now, these studies are yet insufficient to precisely select patients for PC-RPLND to prevent over-treatment [100, 101], inconclusive [99–103] and sometimes controversial [98, 100, 104–106]. Moreover, non-automated approaches for delineation of the regions of interest are time consuming and not repeatable, limiting its practicality in daily practice [107].

microRNAs: a potential target for imaging?

A recent study used molecular analysis to explore a non-imaging method to differentiate between teratoma, viable GCT, and necrosis post-chemotherapy [108]. This approach identified AGR2 and KRT19 as key proteins significantly overexpressed in teratoma compared to necrosis at both microRNA and protein levels.

This approach involved classifying 48 patients into three groups: those with teratoma, those with viable GCT, and those with necrosis. Using a microdissection technique, they precisely isolated representative areas of each tissue type within the lymph nodes [108].

From a nuclear medicine perspective, if these proteins are tagged with a PET tracer, it would shed light on the precise diagnosis in this gray area.

Conclusion

Imaging of NSGCT remains challenging, and while ¹⁸F-FDG PET imaging has limitations, in a few selected scenarios is still able to contribute to clinical management decisions. The experiences with non-¹⁸F-FDG radiotracers have not yet identified a compelling radiotracer for use in this clinical scenario. For now, the complementary benefits of different imaging techniques could be a reasonable approach. The introduction of miRNAs is speculated to revolutionize the field, which are great candidates for future targets to be radiolabeled for imaging NSGCT. The evolving role of radiomics, which remains inconclusive in the field of NSGCT, is still in its infancy but may eventually become a part of routine practice.

Abbreviations

ceCT	contrast-enhanced CT
FAPI	Fibroblast activation protein inhibitor
GPCR	G protein-coupled receptor
iPSCs	induced pluripotent stem cells
LNMRI	lymphotropic nanoparticle MRI
NSGCT	non-seminomatous germ cell tumors
PC-RPLND	Post-chemotherapy retroperitoneal lymph node dissection
PSMA	Prostate-Specific Membrane Antigen
RAID	radioimmunodetection

Acknowledgements

Not applicable.

Author contributions

NA & AMS: Conceptualization NA, EA and AMS: Methodology. EA, MF, MS, AA and HR: Data Collection and initial drafting. NA, AMS and EA: Review & Editing MF and MS: Revision NA and AMS: Supervision * NA and EA contributed equally as first authors.

Funding

None.

Data availability

No datasets were generated or analysed during the current study.

Declarations

Ethics approval and consent to participate

Not applicable. This is a review article.

Consent for publication

All authors have reviewed the final version and consented for publication of the study.

Competing interests

The authors declare no competing interests.

Received: 11 August 2024 / Accepted: 24 October 2024

Published online: 18 November 2024

References

- Kaplan İ, Can C, Güzel Y, Alabalık U, Kömek H. 68 GA-FAPI-04 PET/CT Versus 18 F-FDG PET/CT in imaging of malignant mixed germ cell testicular tumor. *Clin Nucl Med*. 2023;48(4):e195–7.
- Siegel RL, Miller KD, Wagle NS, Jemal A. Cancer statistics, 2023. *CA Cancer J Clin*. 2023;73(1):17–48.
- Nallu A, Mannuel HD, Hussain A. Testicular germ cell tumors: biology and clinical update. *Curr Opin Oncol*. 2013;25(3):266–72.
- Daneshmand S, Albers P, Fosså SD, Heidenreich A, Kollmannsberger C, Krege S, et al. Contemporary management of postchemotherapy testis cancer. *Eur Urol*. 2012;62(5):867–76.
- Nauman M, Leslie SW. Nonseminomatous Testicular Tumors. *StatPearls*. Treasure Island (FL): StatPearls Publishing Copyright © 2024. StatPearls Publishing LLC.; 2024.
- Masterson TA, Tagawa ST. A 25-year review of advances in testicular cancer: Perspectives on evaluation, treatment, and future directions/challenges. *Urol Oncol*. 2021;39(9):561–8.
- Giannopoulou CD. PET/CT Findings in Testicular Cancer. In: Andreou JA, Kosmidis PA, Gouliamos AD, editors. *Artificial Intelligence in PET/CT Oncologic Imaging*. Cham: Springer International Publishing; 2022. pp. 93–8.
- Wakileh GA, Ruf C, Heidenreich A, Dieckmann KP, Lissou C, Prasad V, et al. Contemporary options and future perspectives: three examples highlighting the challenges in testicular cancer imaging. *World J Urol*. 2022;40(2):307–15.
- Oldenburg J, Berney DM, Bokemeyer C, Climent MA, Daugaard G, Gietema JA, et al. Testicular seminoma and non-seminoma: ESMO-EURACAN Clinical Practice Guideline for diagnosis, treatment and follow-up. *Ann Oncol*. 2022;33(4):362–75.
- Mano R, Becerra MF, Carver BS, Bosl GJ, Motzer RJ, Bajorin DF, et al. Clinical Outcome of Patients with Fibrosis/Necrosis at Post-Chemotherapy Retroperitoneal Lymph Node Dissection for Advanced Germ Cell Tumors. *J Urol*. 2017;197(2):391–7.
- Dusaud M, Malavaud B, Bayoud Y, Sebe P, Hoepffner JL, Salomon L, et al. Post-chemotherapy retroperitoneal teratoma in nonseminomatous germ cell tumors: Do predictive factors exist? Results from a national multicenter study. *J Surg Oncol*. 2016;114(8):992–6.
- Santis MD, Maj-Hes A, Bachner M. Positron Emission Tomography (PET) in Germ Cell Tumors (GCT). In: de la Rosette JJMCH, Manyak MJ, Harisinghani MG, Wijkstra H, editors. *Imaging in Oncological Urology*. London: Springer London; 2009. pp. 305–13.
- Conduit C, Hong W, Martin F, Thomas B, Lawrentschuk N, Goad J, et al. A meta-analysis of clinicopathologic features that predict necrosis or fibrosis at post-chemotherapy retroperitoneal lymph node dissection in individuals receiving treatment for non-seminoma germ cell tumours. *Front Oncol*. 2022;12:931509.
- Nestler T, Paffenholz P, Pfister D, Schoch J, Nini A, Hiester A, et al. Adjunctive Surgery Is Often Without Oncological Benefit at Time of Postchemotherapy Retroperitoneal Lymph Node Dissection. *J Urol*. 2024;211(3):426–35.
- Cotner CE, Hilton S, Mamtani R, Guzzo T, Vaughn DJ. Surveillance of post-chemotherapy subcentimeter residual retroperitoneal mass in metastatic nonseminomatous germ cell tumor: Does how you measure matter? *Urol Oncol*. 2021;39(2):136.e11–e17.
- Antonelli L, Ardizzone D, Tachibana I, Adra N, Cary C, Hugar L, et al. Risk Factors for Relapse in Nonseminomatous Testicular Cancer After Postchemotherapy Retroperitoneal Lymph Node Dissection With Viable Residual Cancer. *J Clin Oncol*. 2023;41(34):5296–305.
- Gilligan TD. Resection of Residual Masses After Chemotherapy for Metastatic Nonseminomatous Germ Cell Tumors in Adolescents and Adults. *J Clin Oncol*. 2023;41(23):3899–904.
- Ravi P, Gray KP, O'Donnell EK, Sweeney CJ. A meta-analysis of patient outcomes with subcentimeter disease after chemotherapy for metastatic non-seminomatous germ cell tumor. *Ann Oncol*. 2014;25(2):331–8.
- McAlpine K, Clark R, Spiess PE, Necchi A, Gage K, Hamilton RJ. The Importance of Repeat Imaging Prior to Treatment Decision-making in Testicular Cancer: Commentary From the Inaugural Global Society of Rare Genitourinary Tumors Summit. *Clin Genitourin Cancer*. 2023;21(3):418.e1–e6.
- Revels JW, Wang SS, Gangadhar K, Ali A, Ali AA, Lee JH. Multimodality Radiological Pictorial Review of Testicular Carcinoma: From Initial Staging to Restaging. *Res Rep Urol*. 2020;12:599–613.
- Yacoub JH, Oto A, Allen BC, Coakley FV, Friedman B, Hartman MS, et al. ACR Appropriateness Criteria Staging of Testicular Malignancy. *J Am Coll Radiol*. 2016;13(10):1203–9.
- Jiao H, Qiu Y, Huang W, Zhang Y, Chen Z, Wang A, Kang L. Alpha-fetoprotein-elevated postpubertal testicular teratoma with retroperitoneal metastasis on (18)F-FDG PET/CT: case report and literature review. *Front Med (Lausanne)*. 2023;10:1269587.
- Schieda N, Oto A, Allen BC, Akin O, Barker SJ, Fulgham PF, et al. ACR Appropriateness Criteria® Staging and Surveillance of Testicular Cancer: 2021 Update. *J Am Coll Radiol*. 2022;19(5):S194–207.
- Malik K, Raja A, Radhakrishnan V, Kathiresan N. A retrospective analysis of patients undergoing postchemotherapy retroperitoneal lymph node dissection and metastasectomy in advanced nonseminomatous germ cell tumors. *Indian J Urol*. 2020;36(2):112–6.
- Murez T, Fléchon A, Branger N, Savoie PH, Rocher L, Camparo P, et al. French AFU Cancer Committee Guidelines - Update 2022–2024: testicular germ cell cancer. *Progrès en Urologie*. 2022;32(15):1066–101.
- Steiner H, Berg B, Stöhr B, Fritzer A, Ramoner R, Aigner F et al. Prediction of Retroperitoneal Histology in Metastatic Nonseminomatous Testicular Cancer Patients after Chemotherapy Based on Clinical and Radiological Parameters. *Curr Urol*. 2010;4(3).
- Steyerberg EW, Keizer HJ, Habbema JD. Prediction models for the histology of residual masses after chemotherapy for metastatic testicular cancer. *ReHiT Study Group*. *Int J Cancer*. 1999;83(6):856–9.
- Aparicio J. On behalf of the Spanish Germ Cell Cancer G. Positron emission tomography (PET) is not indicated in the postchemotherapy evaluation of advanced non-seminomatous testicular germ cell tumors. *Clin Transl Oncol*. 2014;16(5):509–10.
- Wood MJ, Tirumani SH, Sweeney C, Ramaiya NH, Howard SA. Approach to risk stratification in testicular germ cell tumors: a primer for radiologists. *Abdom Imaging*. 2015;40(6):1871–86.
- Patrikidou A, Cazzaniga W, Berney D, Boormans J, de Angst I, Di Nardo D, et al. European Association of Urology Guidelines on Testicular Cancer: 2023 Update. *Eur Urol*. 2023;84(3):289–301.
- Rosenvilde JJ, Pedersen GL, Bandak M, Lauritsen J, Kreiberg M, Wagner T, et al. Oncological outcome and complications of post-chemotherapy retroperitoneal surgery in non-seminomatous germ cell tumours - a systematic review. *Acta Oncol*. 2021;60(6):695–703.
- Timothy Gilligan DWL, Adra N, Aggarwal R, Bagrodia A, Costa D, Drakaki A, Emamekhoo H, Evans C, Feldman DR, Geynisman DM, Graham L, Humphrey P, Levine EG, Amy Luckenbaugh, Benjamin Maughan, Bradley McGregor, Paul Monk, Joel Picus, Soroush Rais-Bahrami, Zachery Reichert, Jean-Claude Rwigema, Philip Saylor, Ankeet Shah, Sumit Shah, Nirmish Singla, Kanishka Sircar, Benjamin A. Teplý, David VanderWeele, David Vaughn, Kosj Yamoah, Ali Zhumkhawala. *Testicular Cancer, Version 2.2020, NCCN Clinical Practice Guidelines in Oncology*. National Comprehensive Cancer Network. 2024.

33. Tandstad T, Ståhl O, Håkansson U, Wahlqvist R, Klepp O, Cavallin-Ståhl E, Cohn-Cedermark G. The SWENOTECA group: a good example of continuous binational and multidisciplinary collaboration for patients with testicular cancer in Sweden and Norway. *Scandinavian J Urol*. 2016;50(1):9–13.
34. Ortiz AFH, Beaujon LJF, Villamizar SYG, López FFF. Magnetic resonance versus computed tomography for the detection of retroperitoneal lymph node metastasis due to testicular cancer: a systematic literature review. *Eur J Radiol Open*. 2021;8:100372.
35. Honecker F, Aparicio J, Berney D, Beyer J, Bokemeyer C, Cathomas R, et al. ESMO Consensus Conference on testicular germ cell cancer: diagnosis, treatment and follow-up. *Ann Oncol*. 2018;29(8):1658–86.
36. Lassen U, Daugaard G, Eigtved A, Højgaard L, Damgaard K, Rørth M. Whole-body FDG-PET in patients with stage I non-seminomatous germ cell tumours. *Eur J Nucl Med Mol Imaging*. 2003;30:396–402.
37. Oechsle K, Hartmann M, Brenner W, Venz S, Weissbach L, Franzius C, et al. [18F] Fluorodeoxyglucose positron emission tomography in nonseminomatous germ cell tumors after chemotherapy: the German multicenter positron emission tomography study group. *J Clin Oncol*. 2008;26(36):5930–5.
38. Makovnik M, Rejleková K, Uhrin I, Mego M, Chovanec M. Intricacies of radiographic assessment in testicular germ cell tumors. *Front Oncol*. 2021;10:587523.
39. Quak E, Kovacs I, Oyen WJ, Van der Graaf WT. FDG-PET/CT in a patient with poor-risk non-seminoma testis with mature teratoma and secondary gliosarcoma: multimodality imaging for guiding multimodality treatment. *Nuclear Med Mol Imaging*. 2015;49:237–40.
40. Alongi P, Evangelista L, Caobelli F, Spallino M, Gianolli L, Midiri M, Picchio M. Diagnostic and prognostic value of 18F-FDG PET/CT in recurrent germinal tumor carcinoma. *Eur J Nucl Med Mol Imaging*. 2018;45:85–94.
41. Aydos U, Tahtaci G, Akdemir ÜÖ, Özet A. 18F-FDG PET/CT for primary staging of patients with testicular germ cell tumors: the predictors of 18F-FDG PET positivity and prognostic value of PET derived metabolic parameters. *Nucl Med Commun*. 2020;41(11):1199–209.
42. Spermon JR, De Geus-Oei LF, Kiemeny LA, Witjes JA, Oyen WJ. The role of (18)fluoro-2-deoxyglucose positron emission tomography in initial staging and re-staging after chemotherapy for testicular germ cell tumours. *BJU Int*. 2002;89(6):549–56.
43. Kollmannsberger C, Oechsle K, Dohmen BM, Pfannenberger A, Bares R, Claussen CD, et al. Prospective comparison of [18F] fluorodeoxyglucose positron emission tomography with conventional assessment by computed tomography scans and serum tumor markers for the evaluation of residual masses in patients with nonseminomatous germ cell carcinoma. *Cancer: Interdisciplinary Int J Am Cancer Soc*. 2002;94(9):2353–62.
44. Karapetis C, Strickland A, Yip D, Steer C, Harper P. Use of fluorodeoxyglucose positron emission tomography scans in patients with advanced germ cell tumour following chemotherapy: single-centre experience with long-term follow up. *Intern Med J*. 2003;33(9–10):427–35.
45. Pfannenberger AC, Oechsle K, Bokemeyer C, Kollmannsberger C, Dohmen BM, Bares R, et al. The role of [18 F] FDG-PET, CT/MRI and tumor marker kinetics in the evaluation of postchemotherapy residual masses in metastatic germ cell tumors—prospects for management. *World J Urol*. 2004;22:132–9.
46. Putra LJ, Lawrentschuk N, Ballok Z, Hannah A, Poon A, Tauro A, et al. 18F-fluorodeoxyglucose positron emission tomography in evaluation of germ cell tumor after chemotherapy. *Urology*. 2004;64(6):1202–7.
47. Tsatalpas P, Beuthien-Baumann B, Kropp J, Manseck A, Tiepolt C, Hakenberg OW, et al. Diagnostic value of 18F-FDG positron emission tomography for detection and treatment control of malignant germ cell tumors. *Urol Int*. 2002;68(3):157–63.
48. De Santis M, Pont J. The role of positron emission tomography in germ cell cancer. *World J Urol*. 2004;22(1):41–6.
49. Ambrosini V, Zucchini G, Nicolini S, Berselli A, Nanni C, Allegri V, et al. 18F-FDG PET/CT impact on testicular tumours clinical management. *Eur J Nucl Med Mol Imaging*. 2014;41(4):668–73.
50. Huddart RA, O'Doherty MJ, Padhani A, Rustin GJ, Mead GM, Joffe JK, et al. 18fluorodeoxyglucose positron emission tomography in the prediction of relapse in patients with high-risk, clinical stage I nonseminomatous germ cell tumors: preliminary report of MRC Trial TE22—the NCRI Testis Tumour Clinical Study Group. *J Clin Oncol*. 2007;25(21):3090–5.
51. de Wit M, Brenner W, Hartmann M, Kotzerke J, Hellwig D, Lehmann J, et al. [18F]-FDG-PET in clinical stage I/II non-seminomatous germ cell tumours: results of the German multicentre trial. *Ann Oncol*. 2008;19(9):1619–23.
52. Chen J, Daneshmand S. Modern Management of Testicular Cancer. In: Daneshmand S, Chan KG, editors. *Genitourinary Cancers*. Cham: Springer International Publishing; 2018. pp. 273–308.
53. Kollmannsberger C, Oechsle K, Dohmen BM, Pfannenberger A, Bares R, Claussen CD, et al. Prospective comparison of [18F]fluorodeoxyglucose positron emission tomography with conventional assessment by computed tomography scans and serum tumor markers for the evaluation of residual masses in patients with nonseminomatous germ cell carcinoma. *Cancer*. 2002;94(9):2353–62.
54. Joice GA, Rowe SP, Gorin MA, Pierorazio PM. Molecular imaging for evaluation of viable testicular cancer nodal metastases. *Curr Urol Rep*. 2018;19:1–7.
55. Sugawara Y, Zasadny KR, Grossman HB, Francis IR, Clarke MF, Wahl RL. Germ Cell Tumor: Differentiation of Viable Tumor, Mature Teratoma, and Necrotic Tissue with FDG PET and Kinetic Modeling. *Radiology*. 1999;211(1):249–56.
56. Sharma P, Jain T, Parida G, Karunanithi S, Patel C, Sharma A, et al. Diagnostic accuracy of integrated 18F-FDG PET/CT for restaging patients with malignant germ cell tumours. *Br J Radiol*. 2014;87(1040):20140263.
57. Buchler T, Simonova K, Fencel P, Jarkovsky J, Abrahamova J. Clinical outcomes of patients with nonseminomatous germ cell tumours and negative postchemotherapy positron emission tomography. *Cancer Invest*. 2012;30(6):487–92.
58. Cook GJ, Sohaib A, Huddart RA, Dearnaley DP, Horwich A, Chua S. The role of 18F-FDG PET/CT in the management of testicular cancers. *Nucl Med Commun*. 2015;36(7):702–8.
59. Feldman DR. State-of-the-Art Management of Germ Cell Tumors. *Am Soc Clin Oncol Educ Book*. 2018;38:319–23.
60. Cabral FC, Krajewski KM, Rosenthal MH, Hirsch MS, Howard SA. Teratoma with malignant transformation: report of three cases and review of the literature. *Clin Imaging*. 2014;38(5):589–93.
61. Yoshida M, Tanaka M, Gomi K, Ohama Y, Kigasawa H, Iwanaka T, Tanaka Y. Malignant steroidogenic tumor arising from sacrococcygeal mature teratoma. *Hum Pathol*. 2011;42(10):1568–72.
62. Kole AC, Hoekstra HJ, Sleijfer DT, Nieweg OE, Koops HS, Vaalburg W. L-[1-carbon-11] tyrosine imaging of metastatic testicular nonseminoma germ-cell tumors. *J Nucl Med*. 1998;39(6):1027–9.
63. Pfannenberger C, Aschoff P, Dittmann H, Mayer F, Reischl G, von Weyhern C, et al. PET/CT with 18F-FLT: does it improve the therapeutic management of metastatic germ cell tumors? *J Nucl Med*. 2010;51(6):845–53.
64. Woldu SL, Meng X, Wong D, Baky F, Margulis V, Xi Y, et al. Performance characteristics of 18F-fluciclovine positron emission tomography/computed tomography prior to retroperitoneal lymph node dissection. *Can Urol Association J*. 2022;16(3):E167.
65. Kinoshita Y, Kuratsukuri K, Landas S, Imaida K, Rovito PM, Wang CY, Haas GP. Expression of prostate-specific membrane antigen in normal and malignant human tissues. *World J Surg*. 2006;30:628–36.
66. Simsek DH, Civan C, Ekenel M, Kuyumcu S, Sanli Y. 177Lu-PSMA therapy for metastatic testicular mixed germ cell tumor. *Clin Nucl Med*. 2021;46(5):415–8.
67. Li Y, Liu Z, Dong C, He P, Liu X, Zhu Z, et al. Noninvasive Detection of Human-Induced Pluripotent Stem Cell (hiPSC)-Derived Teratoma with an Integrin-Targeting Agent 99m Tc-3PRGD2. *Mol imaging biology*. 2013;15:58–67.
68. Małkiewicz B, Świrkosz G, Lewandowski W, Demska K, Szczepaniak Z, Karwacki J, et al. Lymph Node Dissection in Testicular Cancer: The State of the Art and Future Perspectives. *Curr Oncol Rep*. 2024;26(4):318–35.
69. Saha GB. *Physics and radiobiology of nuclear medicine*. Springer Science & Business Media; 2012.
70. Crişan G, Moldovean-Cioroianu NS, Timaru D-G, Andrieş G, Căinap C, Chiş V. Radiopharmaceuticals for PET and SPECT imaging: a literature review over the last decade. *Int J Mol Sci*. 2022;23(9):5023.
71. Ward ZJ, Scott AM, Hricak H, Abdel-Wahab M, Paez D, Lette MM, et al. Estimating the impact of treatment and imaging modalities on 5-year net survival of 11 cancers in 200 countries: a simulation-based analysis. *Lancet Oncol*. 2020;21(8):1077–88.
72. Sauerbrunn BJ, Andrews GA, Hübner KF. Ga-67 citrate imaging in tumors of the genito-urinary tract: report of cooperative study. *J Nucl Med*. 1978;19(5):470–5.
73. Bekerman C, Hoffer PB, Bitran JD. The role of gallium-67 in the clinical evaluation of cancer. *Semin Nucl Med*. 1984;14(4):296–323.
74. Willan BD, Penney H, Castor WR, McGowan DG. The usefulness of gallium-67 citrate scanning in testicular seminoma. *Clin Nucl Med*. 1987;12(10):813–5.
75. Jana S, Blaufox MD. *Nuclear Medicine Studies of the Prostate, Testes, and Bladder*. *Semin Nucl Med*. 2006;36(1):51–72.

76. Ballou B, Levine G, Hakala TR, Solter D. Tumor location detected with radioactively labeled monoclonal antibody and external scintigraphy. *Science*. 1979;206(4420):844–7.
77. Javadpour N, Kim EE, DeLand FH, Salyer JR, Shah U, Goldenberg DM. The role of radioimmunodetection in the management of testicular cancer. *JAMA*. 1981;246(1):45–9.
78. Epenetos A, Hooker G, Durbin H, Bodmer W, Snook D, Begent R, et al. Indium-111 labelled monoclonal antibody to placental alkaline phosphatase in the detection of neoplasms of testis, ovary, and cervix. *Lancet*. 1985;326(8451):350–3.
79. Bentestuen M, Al-Obaydi N, Zacho HD, editors. FAPI-avid nonmalignant PET/CT findings: an expedited systematic review. *Seminars in Nuclear Medicine*. Elsevier; 2023.
80. Treglia G, Muoio B, Roustaei H, Kiamanesh Z, Aryana K, Sadeghi R. Head-to-head comparison of fibroblast activation protein inhibitors (FAPI) radiotracers versus [18F] F-FDG in oncology: a systematic review. *Int J Mol Sci*. 2021;22(20):11192.
81. Roustaei H, Kiamanesh Z, Askari E, Sadeghi R, Aryana K, Treglia G. Could fibroblast activation protein (FAP)-specific radioligands be considered as pan-tumor agents? *Contrast Media & Molecular Imaging*. 2022;2022.
82. Pan Q, Luo Y, Zhang W. Idiopathic retroperitoneal fibrosis with intense uptake of 68Ga-fibroblast activation protein inhibitor and 18F-FDG. *Clin Nucl Med*. 2021;46(2):175–6.
83. Xi Y, Sun L, Che X, Huang X, Liu H, Wang Q, et al. A comparative study of [68Ga] Ga-FAPI-04 PET/MR and [18F] FDG PET/CT in the diagnostic accuracy and resectability prediction of ovarian cancer. *Eur J Nucl Med Mol Imaging*. 2023;50(9):2885–98.
84. Pang Y, Zhao L, Meng T, Xu W, Lin Q, Wu H, et al. PET imaging of fibroblast activation protein in various types of cancer using 68Ga-FAP-2286: comparison with 18F-FDG and 68Ga-FAPI-46 in a single-center, prospective study. *J Nucl Med*. 2023;64(3):386–94.
85. Dai Y, Pang Y, Bao W, Cheng Y, Chen H. Pineal yolk sac tumor producing α -fetoprotein detected by 68Ga-FAPI PET/MRI. *Clin Nucl Med*. 2022;47(1):90–2.
86. Kircher M, Herhaus P, Schottelius M, Buck AK, Werner RA, Wester H-J, et al. CXCR4-directed theranostics in oncology and inflammation. *Ann Nucl Med*. 2018;32:503–11.
87. Zhao H, Guo L, Zhao H, Zhao J, Weng H, Zhao B. CXCR4 over-expression and survival in cancer: a system review and meta-analysis. *Oncotarget*. 2015;6(7):5022.
88. Gilbert D, Chandler I, McIntyre A, Goddard N, Gabe R, Huddart R, Shipley J. Clinical and biological significance of CXCL12 and CXCR4 expression in adult testes and germ cell tumours of adults and adolescents. *J Pathology: J Pathological Soc Great Br Irel*. 2009;217(1):94–102.
89. Mallik S, Qin G, Jia P, Zhao Z. Molecular signatures identified by integrating gene expression and methylation in non-seminoma and seminoma of testicular germ cell tumours. *Epigenetics*. 2021;16(2):162–76.
90. Gaytan F, Barreiro M, Caminos J, Chopin L, Herington A, Morales C, et al. Expression of ghrelin and its functional receptor, the type 1a growth hormone secretagogue receptor, in normal human testis and testicular tumors. *J Clin Endocrinol Metabolism*. 2004;89(1):400–9.
91. Charron CL, Hou J, McFarland MS, Dhanvantari S, Kovacs MS, Luyt LG. Structure–activity study of ghrelin (1–8) resulting in high affinity fluorine-bearing ligands for the ghrelin receptor. *J Med Chem*. 2017;60(17):7256–66.
92. Larsen SKA, Løgager V, Bylov C, Nellemann H, Agerbæk M, Als AB, Pedersen EM. Can whole-body MRI replace CT in management of metastatic testicular cancer? A prospective, non-inferiority study. *J Cancer Res Clin Oncol*. 2023;149(3):1221–30.
93. Sohaib S, Koh D, Barbachano Y, Parikh J, Husband J, Dearnaley D, et al. Prospective assessment of MRI for imaging retroperitoneal metastases from testicular germ cell tumours. *Clin Radiol*. 2009;64(4):362–7.
94. Mosavi F, Laurell A, Ahlström H. Whole body MRI, including diffusion-weighted imaging in follow-up of patients with testicular cancer. *Acta Oncol*. 2015;54(10):1763–9.
95. Małkiewicz B, Świrkosz G, Lewandowski W, Demska K, Szczepaniak Z, Karwacki J et al. Lymph Node Dissection in Testicular Cancer: The State of the Art and Future Perspectives. *Curr Oncol Rep*. 2024:1–18.
96. Will O, Purkayastha S, Chan C, Athanasiou T, Darzi AW, Gedroyc W, Tekkis PP. Diagnostic precision of nanoparticle-enhanced MRI for lymph-node metastases: a meta-analysis. *Lancet Oncol*. 2006;7(1):52–60.
97. Harisinghani MG, Saksena M, Ross RW, Tabatabaei S, Dahl D, McDougal S, Weissleder R. A pilot study of lymphotropic nanoparticle-enhanced magnetic resonance imaging technique in early stage testicular cancer: a new method for noninvasive lymph node evaluation. *Urology*. 2005;66(5):1066–71.
98. Baessler B, Nestler T, Pinto dos Santos D, Paffenholz P, Zeuch V, Pfister D, et al. Radiomics allows for detection of benign and malignant histopathology in patients with metastatic testicular germ cell tumors prior to post-chemotherapy retroperitoneal lymph node dissection. *Eur Radiol*. 2020;30(4):2334–45.
99. King KG, Bhanvadia S, Ghodoussipour S, Hwang D, Varghese B, Cen SY, et al. Distinguishing fibrosis/necrosis from teratoma or viable disease in the retroperitoneum in post-chemotherapy, nonseminomatous testicular germ cell tumor using quantitative CT texture analysis. *American Society of Clinical Oncology*; 2018.
100. Fournier C, Leguillotte C, Leblanc E, Le Deley M-C, Carnot A, Pasquier D, et al. Diagnostic Value of the Texture Analysis Parameters of Retroperitoneal Residual Masses on Computed Tomographic Scan after Chemotherapy in Non-Seminomatous Germ Cell Tumors. *Cancers*. 2023;15(11):2997.
101. Scavuzzo A, Pasini G, Crescio E, Jimenez-Rios MA, Figueroa-Rodriguez P, Comelli A, et al. Radiomics Analyses to Predict Histopathology in Patients with Metastatic Testicular Germ Cell Tumors before Post-Chemotherapy Retroperitoneal Lymph Node Dissection. *J Imaging*. 2023;9(10):213.
102. Venishetty N, Taylor J, Xi Y, Howard JM, Ng YS, Wong D, et al. Testicular radiomics to predict pathology at time of postchemotherapy retroperitoneal lymph node dissection for nonseminomatous germ cell tumor. *Clin Genitourin Cancer*. 2024;22(1):33–7.
103. NITTA S, KOJIMA T, GIDO M, NAKAGAWA S, KAKEYA H, KANDORI S, et al. A Machine Learning Model to Predict the Histology of Retroperitoneal Lymph Node Dissection Specimens. *Anticancer Res*. 2024;44(5):2151–7.
104. King KG, Bhanvadia S, Ghodoussipour S, Hwang D, Varghese B, Cen SY, et al. Distinguishing fibrosis/necrosis from teratoma or viable disease in the retroperitoneum in post-chemotherapy, nonseminomatous testicular germ cell tumor using quantitative CT texture analysis. *J Clin Oncol*. 2018;36(6suppl):563.
105. Lewin J, Dufort P, Halankar J, O'Malley M, Jewett MAS, Hamilton RJ, et al. Applying Radiomics to Predict Pathology of Postchemotherapy Retroperitoneal Nodal Masses in Germ Cell Tumors. *JCO Clin Cancer Inf*. 2018;2:1–12.
106. Nestler T, Baeßler B, Pinto dos Santos D, Pfister DJ, Maintz D, Heidenreich A. Predicting vital retroperitoneal residual tumors of metastatic testicular tumor patients after chemotherapy using radiomics. *American Society of Clinical Oncology*; 2019.
107. Ding R, Li X, Liu Z. Overview of Novel Biomarkers for Management of Post-chemotherapy Residual Masses in Testicular Cancer. *Eur Urol Focus*. 2024.
108. Nestler T, Kremer L, von Brandenstein M, Wittersheim M, Paffenholz P, Wagener-Rydzek S, et al. Differentially expressed messenger RNA/proteins can distinguish teratoma from necrosis in postchemotherapy retroperitoneal lymph node dissection tissue. *Cancer*. 2023;129(4):634–42.
109. Katabathina VS, Vargas-Zapata D, Monge RA, Nazarullah A, Ganeshan D, Tammisetti V, Prasad SR. Testicular Germ Cell Tumors: Classification, Pathologic Features, Imaging Findings, and Management. *Radiographics*. 2021;41(6):1698–716.
110. Aide N, Briand M, Bohn P, Dutoit S, Lasnon C, Chasle J, et al. α v β 3 imaging can accurately distinguish between mature teratoma and necrosis in 18F-FDG-negative residual masses after treatment of non-seminomatous testicular cancer: a preclinical study. *Eur J Nucl Med Mol Imaging*. 2011;38(2):323–33.
111. Cao F, Li Z, Lee A, Liu Z, Chen K, Wang H, et al. Noninvasive De novo Imaging of Human Embryonic Stem Cell–Derived Teratoma Formation. *Cancer Res*. 2009;69(7):2709–13.
112. Bohn P, Modzelewski R, Rouvet J, Briand M, Dutoit S, Pille J-Y et al. Biodistribution and imaging of [99mTc]-HYNIC-RGD in MDA-MB-231 and NTERA-2 cancer cell xenografts. *Nucl Med Commun*. 2013;34(7).
113. Lehner S, Lang C, Kaisis G, Todica A, Zacherl MJ, Boening G, et al. 124 I-PET assessment of human sodium iodide symporter reporter gene activity for highly sensitive in vivo monitoring of teratoma formation in mice. *Mol imaging biology*. 2015;17:874–83.
114. Stroosma OB, Delaere KP. Carcinoid tumours of the testis. *BJU Int*. 2008;101(9):1101–5.
115. Amine MM, Mohamed B, Mourad H, Majed H, Slim C, Mehdi B, et al. Neuroendocrine Testicular Tumors: A Systematic Review and Meta-Analysis. *Curr Urol*. 2017;10(1):15–25.

116. Center UJCC. Prospective Exploratory Study of FAPI PET/CT with Histopathology Validation in Patients with Various Cancers. ClinicalTrialsgov IDNCT04459273.

Publisher's note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.